

Evaluation of some horticultural traits and resistance to Fusarium wilt of some watermelon hybrids

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Abstract— Fusarium wilt is a major disease of watermelon in many parts of the world. Control of this disease is difficult because the soil borne causal agent *Fusarium oxysporum* f. sp. *niveum* (Fon) produces chlamydospores that remain infectious in the soil for many years. In this study, four lines, three testers of watermelon and their 12 F₁ hybrids which were developed through line X tester mating design were used to study the mean performance and heterosis for six horticultural traits and resistance to Fusarium wilt. Biochemical parameter such as phenolic content and three oxidative-reductive enzymes (PAL, PPO and PO) in three watermelon hybrids shown susceptible, moderately resistant and resistant to the fungus *F.o.* f.sp. *niveum* were assessed. All parents and crosses were significant with positive value for all traits studied. With respect to heterosis effects, cross P₂xP₆ gave the highest values with significant effect for plant fresh weight, total yield/plant and fruit weight for mid-parent and better parent. Crosses P₂xP₇, P₃xP₅ and P₄xP₆ gave earliest female flower over mid-parent and better parent. In addition, cross P₄xP₆ gave the highest values with significant effect for rind thickness for mid-parent and better parent. Moreover, cross P₁xP₇ gave the highest values with significant effect for T.S.S. for mid-parent. Results showed that the lowest mean performance of seven parents and their twelve crosses of watermelon for the average of the severity of Fusarium wilt on the foliage growth and on the xylem vesicles were 21.9 % for parents line (Beni-Swif ; P₇) and 26.2 % for crosses (P₂xP₇). Also, data revealed that, in general, the three assessed enzymes, i.e. PAL, PPO and PO and total phenols were greatly increased in the shoot base of the hybrids by increasing the resistance of these hybrids.

Key words: Watermelon, lines, hybrids, mean performance, heterosis, Fusarium wilt, total phenols, oxidative-reductive enzymes.

1 INTRODUCTION

Watermelon, (*Citrullus lanatus*, Thunp.), is one of the most popular vegetable crops, especially in summer season in Egypt. All over the world, many investigators are interested in collecting their landraces or accession lines and estimate their agronomic traits or pathogenicity test in order to raise a new genotype for breeding programs (Dane *et al.*, 1998 and Khereba *et al.*, 2007

Fusarium wilt [caused by *Fusarium oxysporum* f. sp. *niveum* (Fon)] is one the most serious soil-borne diseases of water-melon (Netzer, 1976 ; Martyn, 1985 and 1987 and Mahdy *et al.*, 2014). Initial symptoms can vary with chlorosis of leaves being common followed by asymmetrical wilting for the plant. When infection takes place in seedling, noticeable stunting of the plant occurs followed by chlorosis and death. Necrosis of the stem, in the form of brown streaks in the vascular system produced by both fungal exudates as well as the dying plant tissues, are characteristic of Fusarium wilt (FW).

To date, the disease is best controlled through the use of wilt-resistant cultivars and crop rotations for a minimum of 5 to 7 years (Hopkins and Elmstrom, 1984). There are also opinions that the best solution is to develop highly tolerant cultivars (Martyn, and McLaughlin, 1983). In order to test the resistance of new breeding material, it is necessary to have reliable screening methods.

Two U.S. PIs of *Citrullus Lanatus* var. *citroides* (PI 296341 and PI 271769) have been reported as resistant to Race- 2 of Fon (Dane *et al.*, 1998; Martyn and Netzer, 1991).

Survey 110 U.S. PIs of wild watermelon (*Citrullus Lanatus*

var. *citroides*) for resistance to Race-2 (FW). Of these 110 accessions, 15 showed significantly higher resistance to Fon Race-2 than that found in the watermelon cultivars Sugar Baby or Charleston Grey as well as in the *C. Lanatus* var. *citroides* PI 296341 that was reported to contain resistance to FW. Wechter *et al.* (2012) studied the genetic behavior of some economic characters in watermelon. They found that all studied traits were influenced by both the additive and non-additive gene action. Some traits (number of branches, number of fruits/plant and T.S.S.) showed partial dominance. Meanwhile, four traits (leaves number, days to female flowering, total yield and fruit length) showed complete dominance. The stem length, average fruit weight and number of seeds/fruit showed over-dominance (El-Maghawry *et al.* 2001)

The aim of this research was to evaluate the level of resistance to *F. oxysporum* f. sp. *niveum* in selected inbred lines and hybrids of watermelon. In order to establish a breeding program for watermelon to find resistance source and produce some watermelon F₁ hybrids, which many give high yield in quantity and quality with resistance to Fusarium wilt.

2 MATERIALS AND METHODS

The experiment was conducted at Kaha Vegetable Research Farm (KVRF), Qalubia governorate, Egypt. The material for this study presented in Table (1) was comprised between seven genotypes. In summer season (2015) the best 4 inbred lines, based on evaluation of resistance to Fusarium wilt for lines (Suhag-1, Suhag-2. Sinaa, Bir El-Abd-2) were used as female

parents; and the cultivars Giza-1, Crimson sweet and the line (Beni-Swif) were used as male parents. Their 12 crosses were developed through line x tester- mating design.

2.1. Evaluation of horticultural characters for parents and hybrids :

| Parents | Skin color | Flesh color | Fruit shape |
|--------------------|-------------------------------|-------------|-------------|
| Suhag1 (P1) | Green striped with dark green | red | Round |
| Suhag2 (P2) | Light green | yellow | Round |
| Sinaa (P3) | dark green | red | Round |
| Bir El-abd2 (P4) | green | yellow | Round |
| Giza1 (P5) | green | red | Round |
| Crimson sweet (P6) | Green striped with dark green | rose | Round |
| Beni Swif (P7) | dark green | yellow | Round |

In summer season (2016) , seeds of the seven parents (lines and testers) and twelve hybrids were planted at KVERF. The experimental design used was a randomized complete block design (RCBD) with three replicates.

Standard cultural practices were adopted as recommended by the Min. of Agric and Land Recl. Data were recorded for some characters, i.e. lant fresh weight, number of days to flowering, total yield per plant, fruit weight, rind thickness and total soluble solids (T.S.S.).

TABLE 1.

NAMES OF PARENTS AND DESCRIPING

2.2. Statistical analysis:

Analyses of variance were carried out using computer software program. The differences among means for all traits were tested for significance using the method described by Snedecor and Cochran (1980).

Estimates of heterosis based on mid parental value and better parent (BP) were determined for each F₁ hybrid according to the following equation (Sinha and Khanna, 1975)

$$\text{Mid-parent heterosis} = \frac{\overline{F_1} - \overline{MP}}{\overline{MP}} \times 100$$

$$\text{Heterobeltiosis (BP)} = \frac{\overline{F_1} - \overline{BP}}{\overline{BP}} \times 100$$

Where: F₁ = Mean performance of the F₁ hybrid; MP = Mean performance of P₁ and P₂.
BP = Mean performance of the better parent

2.3. Evaluation of resistance to Fusarium wilt for parents and hybrids:

Evaluation of parents and hybrids was in summer season (2016) against infection with *F.o. f.sp. niveum*, the causal fungus isolate obtained from Mycol. Res. and Dis.Sur.Dept., Plant Pathol. Res. Instit., ARC.

In summer season (2016) five plants were planted in each plastic pot (35 cm. in diameter), three replicate for each line serve as treatment and three others serve as control, after one month of planting treatment replicate inoculated with spore suspension of the causal fungus (1X10⁶ conidia/ml water) at the rate of 50 ml/pot (Abada and Ahmed, 2014). The plants were examined for severity of infection by the tested fungus one month after inoculation. The pots were irrigated when it was necessary and fertilized with recommended doses by Min. of Agric. and Land Recl.

2.4. Disease assessment:

Disease severity was assessed one month after inoculating watermelon seedlings of one month old using the devised scale (0 to 5) by Amini and Sidovich (2010) on the foliage growth using the following scale:

Where:

0 = No foliar symptoms.

1 = Chlorosis and/or wilt restricted to the first leaf.

2 = Chlorosis and/or wilt extending beyond the first leaf.

3 = Moderate to severe foliar symptoms usually with some absent leaves.

4 = Severe foliar symptoms on the entire plant.

5 = Dead plant.

Disease severity on foliage growth % = $\sum (nxv)/5N \times 100$

Where:

n = Number of infected leaves in each category.

V = Numerical values of each category.

N = Total number of the infected leaves.

The plants were also rated for vascular discoloration using the devised scale (0-5) by Ulloa *et al.* (2006) using the following scale:

Where:

0 = No discoloration.

1 = Light discoloration evident as spotty areas in the longitudinal section of the basal stem and the crown.

2= More continuous discoloration covering an area between one quarter and one half of the longitudinal section of the basal stem and the crown but with light in color.

3= Vascular discoloration (moderate in color) evident in a band encircling almost the entire basal stem and the crown.

4 = Vascular discoloration darker in color than in 1 or 2, and evident across most of the vascular tissue in the longitudinal section of the basal stem and the crown.

5= Plant severely damaged, vascular discoloration evident throughout the longitudinal-section of the basal stem and the crown.

Disease severity on the vascular % = $\sum (nxv)/5N \times 100$

Where:

n = Number of infected vascular in each category.

V = Numerical values of each category.

N = Total number of the infected vascular.

2.4. Determination of the changes in the activity of oxidative-reductive enzymes in three hybrids shown susceptible, moderately resistance and resistant to the causal fungus :

Determination of phenylalanine ammonia lyase (PAL) activity was determined in Laboratory analysis and measurement, Central Laboratory for Biotechnol. of Agric. Res. Cent., Plant Pathol. Res. Instit.; according to the method described by Zucker (1965). The determination of polyphenol oxidase (PPO) activity according to the method described by Esterbaner *et al.* (1977). Determination of peroxidase (PO) activity was determined according to the method described by Worthington (1972).

2.5. Determination of free, conjugated and total phenols in three watermelon hybrids shown susceptible, moderately resistance and resistant to the causal fungus:

Free, conjugated and total phenols were determined in Laboratory analysis and measurement, Central Laboratory for Biotechnol. of Agric. Res. Cent., Plant Pathol. Res. Instit. according to the method described by Simons and Ross (1971).

3 RESULTS AND DISCUSSION

3.1. Evaluation of horticultural characters:

3.1.1. Mean performance:

a. Plant fresh weight:

Plant fresh weight indicated highly significant differences among parents and their twelve crosses as in Table (2). It ranged from 0.802 Kg/plant (Beni-Suif) to 1.163 Kg/plant ($P_2 \times P_6$). The ecotype Sohag-1 recorded the highest values in parents, while the highest values in Crosses ($P_2 \times P_6$). The obtained results are in agreement with those reported by El-Shoura (2007).

b. Number of days to first female flower anthesis:

Data presented in Table (2) indicate that, the parents, Sohag-1 and the hybrid $P_3 \times P_5$ gave the lowest values of NDF; this indicated the earliness of these parents and crosses. Similar results were obtained by Rajan *et al.* (2002) and El-Maghawry *et al.* (2001).

c. Total yield/plant (k.g):

Data presented in Table (2) reveal that, the parents, Sohag-1 and the hybrid $P_1 \times P_6$ gave the highest values of total yield/plant. These results are in agreement with those re-

ported by Rajan *et al.* (2002) and Gilli and Kumai (1988).

d. Fruit weight (kg):

Fruit weight indicated highly significant differences among parents and their twelve crosses as shown in Table (3). It ranged from 2.47 Kg ($P_3 \times P_7$) to 4.29 Kg ($P_1 \times P_6$). The ecotype Sohag-1 recorded the heaviest values in parents, while the heaviest values were found in crosses ($P_1 \times P_6$). Similar results were obtained by El-Maghawry *et al.* (2001).

e. Rind thickness:

Data presented in Table (3) show that, the parents, Giz-1 and the hybrid $P_4 \times P_6$ gave the highest values of rind thickness. These results are in agreement with those reported by Khoreba *et al.* (2007).

f. Total soluble solids (T.S.S.):

T.S.S. indicated highly significant differences among parents and their twelve crosses as shown in Table (3). The ecotype Giza-1 recorded the highest values in parents, while the highest value was found in crosses ($P_1 \times P_7$). Similar results were obtained by El-Maghawry *et al.* (2001) and Khoreba *et al.* (2007).

3.1.2. Heterosis:

a. Plant fresh weight (kg):

Obtained heterosis values based on mid-parent that presented in Table (4) show that, nine F_1 hybrids were significantly differed in plant fresh weight. Estimated heterosis percentages relative to better parent (B.P.) showed that, five crosses showed significant positive values, suggesting over-dominance for large parent. This result is in agreement with those reported by El-Shoura (2007)

b. Number of days to first female flower anthesis:

The estimated heterosis values based on mid-parent are presented in Table (4). Data show that, seven F_1 hybrids gave significant negative differed in plant fresh weight, suggesting dominance towards the short period, since they significantly decreased number of days to first female flower anthesis. Estimated heterosis percentages relative to better parent showed that, three crosses showed significant negative values, suggesting over-dominance for short period. These results are in agreement with those reported by Rajan *et al.* (2002).

TABLE 2

MEAN PERFORMANCE OF SEVEN PARENTS AND THEIR TWELVE CROSSES OF WATERMELON FOR PLANT FRESH WEIGHT, NUMBER OF DAYS TO FIRST FEMALE FLOWER ANTHESIS AND TOTAL YIELD /PLANT

| Genotypes | Plant fresh weight | No. of days | |
|-------------------|--------------------|---------------------------------|-------------------------|
| | | to first female flower anthesis | Total yield/plant (k.g) |
| Sohag 1 (P1) | 1.097 | 47.83 | 8.42 |
| Sohag 2 (P2) | 0.964 | 56.50 | 6.87 |
| Sinaa (P3) | 0.928 | 52.13 | 6.30 |
| Bir El- abd2(P4) | 0.934 | 58.73 | 7.07 |
| Giza1(P5) | 0.907 | 54.73 | 6.70 |
| Crimson sweet(P6) | 0.873 | 63.46 | 6.56 |
| Beni Swif (P7) | 0.802 | 56.53 | 6.79 |
| P1xP5 | 1.050 | 50.76 | 8.24 |
| P1xP6 | 1.113 | 55.73 | 9.02 |
| P1xP7 | 0.923 | 54.06 | 8.24 |
| P2xP5 | 1.050 | 58.46 | 6.56 |
| P2xP6 | 1.163 | 62.40 | 8.43 |
| P2xP7 | 1.063 | 53.80 | 7.30 |
| P3xP5 | 1.087 | 49.90 | 6.52 |
| P3xP6 | 0.860 | 56.06 | 6.19 |
| P3xP7 | 0.960 | 58.20 | 5.72 |
| P4xP5 | 1.047 | 55.03 | 6.15 |
| P4xP6 | 0.977 | 57.30 | 8.21 |
| P4xP7 | 0.937 | 59.56 | 6.57 |
| LS.D (0.05) | 0.061 | 0.903 | 0.40 |
| LS.D (0.01) | 0.087 | 1.29 | 0.58 |

c. Total yield/plant (k.g):

Results shown in Table (4) show that, heterosis values from mid-parents (M.P.) showed significant values for the six crosses. Estimated heterosis percentages relative to better parent showed that, four crosses showed significant positive values, suggesting over-dominance for large parent. Similar results were obtained by Rajan *et al.* (2002).

TABLE 3

MEAN PERFORMANCE OF SEVEN PARENTS AND THEIR TWELVE CROSSES OF WATERMELON FOR FRUIT WEIGHT, RIND THICKNESS AND TOTAL SOLUBLE SOLIDS (TSS)

| Genotypes | Fruit weight (kg) | rind thickness | TSS |
|-------------------|-------------------|----------------|-------|
| Sohag 1 (P1) | 4.24 | 1.38 | 8.53 |
| Sohag 2 (P2) | 3.49 | 1.28 | 8.93 |
| Sinaa (P3) | 2.74 | 1.10 | 8.50 |
| Bir El- abd2(P4) | 3.55 | 1.24 | 9.40 |
| Giza1(P5) | 3.14 | 1.53 | 10.16 |
| Crimson sweet(P6) | 3.14 | 0.94 | 9.10 |
| Beni Swif (P7) | 3.47 | 1.29 | 9.53 |
| P1xP5 | 4.18 | 1.52 | 9.37 |
| P1xP6 | 4.29 | 1.55 | 9.27 |
| P1xP7 | 4.01 | 1.36 | 9.70 |
| P2xP5 | 2.82 | 1.61 | 9.50 |
| P2xP6 | 4.20 | 1.23 | 9.40 |
| P2xP7 | 3.38 | 1.29 | 9.47 |
| P3xP5 | 2.79 | 0.95 | 8.70 |
| P3xP6 | 2.60 | 0.92 | 8.77 |
| P3xP7 | 2.47 | 1.42 | 8.90 |
| P4xP5 | 2.74 | 0.97 | 9.50 |
| P4xP6 | 3.75 | 1.68 | 8.80 |
| P4xP7 | 2.89 | 1.14 | 9.40 |
| LS.D (0.05) | 0.28 | 0.061 | 0.168 |
| LS.D (0.01) | 0.41 | 0.087 | 0.242 |

TABLE 4

Average degree of heterosis (ADH) % based on mid-parent (M.P.) and better-parent of 12 F₁ hybrids for some watermelon characters

| Crosses | Plant fresh weight (kg) | | Number of days to first female flower anthesis | | Total yield /plant (kg) | |
|---------|-------------------------|---------|--|---------|-------------------------|---------|
| | MP | BP | MP | BP | MP | BP |
| | P1xP5 | 5.21 | -3.67 | -1.01** | 6.21 | 9.01** |
| P1xP6 | 13.37** | 2.14 | 0.15** | 16.60 | 20.42** | 7.38** |
| P1xP7 | -2.39 | -15.29 | 3.67 | 13.11 | 8.65** | -1.83 |
| P2xP5 | 12.17** | 8.92* | -2.72** | 6.89 | -3.38 | -4.51 |
| P2xP6 | 26.58** | 20.67** | 4.0 | 10.44 | 25.44** | 22.70** |
| P2xP7 | 20.42** | 10.30** | -4.78** | -4.78** | 6.93* | 6.30* |
| P3xP5 | 18.37** | 17.09** | -6.55** | -4.22** | 0.15 | -2.83 |
| P3xP6 | -4.55 | -7.32 | -2.99** | 7.61 | -3.88 | -5.78 |
| P3xP7 | 10.98** | 3.44 | 7.18 | 11.71 | -12.62 | -15.70 |
| P4xP5 | 13.64** | 12.06** | -2.93** | 0.61** | -10.64 | -13.04 |
| P4xP6 | 8.03* | 4.56 | -6.22** | -2.39** | 20.47** | 16.05** |
| P4xP7 | 7.91* | 0.28 | 3.41 | 5.43 | -5.33 | -7.20 |

Sohag 1 (P1), Sohag 2 (P2), Sinaa (P3), Bir El- abd2(P4), Giza1(P5), Crimson sweet(P6), Beni Swif (P7)

***: significant and highly significant at 0.05 and 0.01 level, respectively

Mp: mid-parent Bp: Better parent

d.

Fruit weight (Kg):

The obtained heterosis values based on mid-parent are presented in Table (5). Results show that, four F₁ hybrids were significantly differed in fruit weight. Estimated heterosis percentages relative to better parent showed that, two crosses showed significant positive values, suggesting over-dominance for large parent. Similar results were obtained by El-Maghawry *et al.* (2001) and Khereba *et al.* (2007).

e. Rind thickness:

Estimates of heterosis values based on mid-parent are presented in Table (5). Results show that, six F₁ hybrids gave significant differed in rind thickness. Estimated heterosis percentages relative to better parent showed that, four crosses showed significant values, suggesting over-dominance towards larger parent. These results are in agreement with those found by Khereba *et al.* (2007).

f. Total soluble solids (T.S.S.):

Results presented in Table (5) reveal that, heterosis values from mid-parents (M.P.) showed significant values four crosses. Estimated heterosis percentages relative to better parent showed that, three crosses showed significant positive values, suggesting over-dominance for large parent. Similar results were obtained by El-Maghawry *et al.* (2001) and Khereba *et al.* (2007).

TABLE 5

Average degree of heterosis (ADH) % based on mid-parent (M.P.) and better-parent of 12 F₁ hybrids for some watermelon characters

| Crosses | Fruit weight (kg) | | Rind thickness | | T.S.S | |
|---------|-------------------|---------|----------------|---------|--------|--------|
| | MP | BP | MP | BP | MP | BP |
| P1xP5 | 13.36** | -1.34 | 4.47* | -0.44 | 0.21 | -7.90 |
| P1xP6 | 16.26** | 1.18 | 33.62** | 12.08** | 5.10** | 1.83* |
| P1xP7 | 4.15 | -5.42 | 1.87 | -1.21 | 7.42** | 1.78* |
| P2xP5 | -14.93 | -19.20 | 14.59** | 5.23* | -0.52 | -6.59 |
| P2xP6 | 26.69** | 20.34** | 10.81** | -3.64 | 4.27** | 3.29** |
| P2xP7 | -2.87 | -3.15 | 0.389 | 0.25 | 2.60** | -0.66 |
| P3xP5 | -4.98 | -11.04 | -27.75 | -38.12 | -6.85 | -14.45 |
| P3xP6 | -11.56 | -17.19 | -9.80 | -16.36 | -0.34 | -3.66 |
| P3xP7 | -20.45 | -28.81 | 18.82** | 10.07** | -1.27 | -6.61 |
| P4xP5 | -17.98 | -22.72 | -29.96 | -36.8 | -2.96 | -6.58 |
| P4xP6 | 12.20** | 5.72* | 54.12** | 35.21** | -4.86 | -6.38 |
| P4xP7 | -17.66 | -18.59 | -9.88 | -11.36 | -0.63 | -1.36 |

Sohag 1 (P1), Sohag 2 (P2), Sinaa (P3), Bir El- abd2(P4), Giza1(P5), Crimson sweet(P6), Beni Swif (P7)

*, **: significant and highly significant at 0.05 and 0.01 level, respectively
mid-parent Bp: Better parent

3.2. Evaluation of resistance to Fusarium wilt for parents and hybrids:

Fusarium wilt of watermelon is a soilborne disease. The fungus attacks seed, seedlings, and roots. The fungus spreads from roots through the vascular system as microconidia reaching all parts of the plant. The fungus is persistent in the soil and is spread long distances on infected seed. Once a host is infected, the fungus increases dramatically. Both macroconidia and chlamydospores are produced on the surface of tissue killed by the organism.

Data shown in Table (6) indicate that the performance of seven parents and their twelve crosses of watermelon for disease severity on the foliage growth were ranged between 24.3-48.9% for crosses (P₄xP₅) and (P₁xP₅); Respectively. The highest percentages of disease severity on the foliage growth were 24.8, 27.0 and 29.5% for parents P₇, P₄ and P₅, respectively. Meanwhile, the lowest percentages of disease severity on the foliage growth were 24.3, 24.4, and 34.7 % for crosses (P₄xP₅), (P₂xP₇) and (P₄xP₇), respectively. Meanwhile, the performance of seven parents and their twelve crosses of watermelon for disease severity on the xylem vesicles were ranged between 14.30- 60% for parent P₆ and crosses (P₁xP₅), respectively. In addition, the lowest percentages of disease severity on the xylem vesicles were 14.30, 19.0, and 27.0% for parents P₆, P₇ and P₄, respectively. In addition, the lowest percentages disease severity on the xylem vesicles were 28, 28.8, and 30% for crosses (P₄xP₇), (P₂xP₇) and (P₄xP₅), respectively. Meanwhile, parents P₇, P₆ and P₄ and crosses (P₂xP₇), (P₄xP₅), (P₄xP₇), (P₂xP₅) and (P₃xP₇) showed the lowest percentages of disease severity on the average, respectively. From these data we can classified the plants according to

the disease resistance : resistant plant showed 20-30% disease severity in the average, moderately resistance plant showed >30-40% disease severity in the average and the susceptible plant showed >40% disease severity in average. (Elmstrom and Hopkins 1981 and Swiader *et al* 2002)

TABLE 6

Mean performance of seven parents and their twelve crosses of watermelon for disease severity on the foliage growth and xylem vesicles

| Treatments | Pal mg/g fw | PPO activity mg/g fw | PO activity mg/g fw | Genotype |
|---------------------------------|----------------|-------------------------|------------------------|----------|
| | | | | |
| susceptible treatment | 0.0151 | 0.0215 | 0.0493 | |
| P2xP5 control | 0.0429 | 0.0273 | 0.0548 | |
| moderately resistance treatment | 0.0548 | 0.0277 | 0.0765 | |
| P2xP7 control | 0.0483 | 0.0311 | 0.1107 | |
| resistant treatment | 0.0765 | 0.0550 | 0.1840 | |
| Sohag 1 (P1) | | | | |
| Sohag 2 (P2) | 43.80 | 31.70 | 37.75 | -- |
| Sinaa (P3) | 44.20 | 47.80 | 45 | -- |
| Bir El- abd2 (P4) | 27.0 | 27.00 | 27 | R |
| Giza1 (P5) | 29.50 | 29.40 | 29.45 | -- |
| Crimson sweet (P6) | 38.30 | 14.30 | 26.30 | R |
| Beni Swif (P7) | 24.80 | 19.00 | 21.9 | R |
| P1xP5 | 48.90 | 60.00 | 54.45 | S |
| P1xP6 | 43.20 | 57.00 | 50.1 | S |
| P1xP7 | 46.20 | 38.50 | 42.35 | -- |
| P2xP5 | 40.00 | 36.00 | 38.0 | mR |
| P2xP6 | 42.70 | 56.70 | 49.7 | -- |
| P2xP7 | 24.40 | 28.80 | 26.2 | R |
| P3xP5 | 40.70 | 44.30 | 42.5 | -- |
| P3xP6 | 38.60 | 41.90 | 40.25 | -- |
| P3xP7 | 36.30 | 40.00 | 38.15 | mR |
| P4xP5 | 24.30 | 30.00 | 27.15 | R |
| P4xP6 | 38.30 | 41.80 | 40.05 | -- |
| P4xP7 | 34.70 | 28.00 | 31.35 | mR |
| L.S.D (0.05) | 3.09 | 2.02 | ---- | |
| L.S.D (0.01) | 4.45 | 2.91 | ----- | |

S; susceptible, mR; moderately resistance and R; resistant.

3.3. Determination of changes in the activity of oxidative-reductive enzymes in three hybrids shown susceptible, medium resistance and resistant to the causal fungus:

Table (7) shows the changes in the activity of oxidative-reductive enzymes, *i.e.* phenylalanine ammonia lyase (PAL), polyphenol oxidase (PPO) and peroxidase (PO) in different hybrids (P₁xP₅, P₂xP₅, P₂xP₇) shown susceptible, moderately resistance and resistant to fungus *fusarium oxysporum* f.sp. *niveum*, respectively. Data reveal that, in general, the three enzymes, *i.e.* PAL, PO and PPO were greatly increased in the shoot base of the hybrids by increasing the resistance of these hybrids. The three enzymes, *i.e.* PAL, PO and PPO were greatly increased in plant treated with fusarium than control plants. It is well known that PAL is a key enzyme of phenylpropanoid metabolism which leads to the synthesis of phenols (Massala *et al.* 1980 and Mahdy *et al.* 2014). PO is a key enzyme in the biosynthesis of lignin and other oxidised phenols (Bruce & West 1989). PO catalyzes the oxidation of hydroxy cinnamyl alcohols into free radical intermediates, which subsequently are coupled into lignin polymers (Gross, 1980). It is

well known that PPO is a copper containing enzyme, which is responsible for oxidization of phenolics to highly toxic quinines. This enzyme is also involved in terminal oxidation of diseased plant tissue, and this role of this enzyme is attributed in disease resistance (Kosuge, 1996).

TABLE 7

Determination of changes in the activity of oxidative-reductive enzymes in three hybrids shown susceptible, medium resistance and resistant to the causal fungus

3.4. Determination of free, conjugated and total phenols in three watermelon hybrids shown susceptible, moderately resistance and resistant to the causal fungus:

Table (8) shows the amount of free, conjugate and total phenols in three watermelon hybrids shown susceptible, moderately resistance and resistant to Fusarium wilt. Free phenols, being 4.745, 5.343 and 5.443 for susceptible, moderately resistance and resistant plants infected by FW, respectively. and being 4.235, 4.839 and 4.771 for control plants. Meanwhile, conjugated phenols being 2.163, 3.120 and 4.478 for susceptible, moderately resistance and resistant plants infected by FW, respectively. and being 2.070, 2.286 and 3.810 for control plants, respectively. Total phenols being 6.908, 8.463 and 9.921 for susceptible, moderately resistance and resistant plants infected with FW, respectively. and being 6.305, 7.125 and 8.581 for control plants. In general the amount of free phenols was more than conjugated phenols in all infected plants with FW and control plants. Also, total phenols in infected plants were more than in the control and increase in resistant plants than susceptible plants.

Phenolic compounds play three main roles in resistance against pathogens; direct attacks on pathogens through antimicrobial compounds, defence of plant cell walls as physical barriers, and induction of resistant responses as molecular messengers (Nicholson and Hammerschmidt 1992). It is well known that phenolic compounds are fungi toxic in nature. Moreover, phenolic compounds increase the physical and mechanical strength of host cell wall resulting in inhibition or fungal invasion (Anand *et al.*, 2007).

TABLE 8

Values of free, conjugated and total phenols in three watermelon hybrids shown susceptible, moderately resistance and resistant to the causal fungus

| Treatments | | Amount of phenolic compounds as mg/g fresh weight | | |
|-----------------------------------|-----------|---|------------|-------|
| | | Free | Conjugated | Total |
| P1xP5 susceptible | control | 4.235 | 2.070 | 6.305 |
| | treatment | 4.745 | 2.163 | 6.908 |
| P2xP5 moderately resistance | control | 4.839 | 2.286 | 7.125 |
| | treatment | 5.343 | 3.120 | 8.463 |
| P2xP7 resistant | control | 4.771 | 3.810 | 8.581 |
| | treatment | 5.443 | 4.478 | 9.921 |

4 REFERENCES

- [1] Abada, K.A. and M.A. Ahmed (2014). Management Fusarium wilt of sweet pepper by Bacillus strains. The American J. of Life Sci., 2(3): 19-25.
- [2] Anand, T. ; A.Chandrasekaran ; S. Kuttalam ; T. Raguchander ; V. Prakasam and R. Samiyappan (2007) Association of some plant defense enzyme activities with systemic resistance to early leaf blight and leaf spot induced in tomato plants by Azoxystrobin and Pseudomonas fluorescens. J. of Plant Interac., 2, 233-244.
- [3] Amini, J. and D.F. Sidovich (2010). The effects of fungicides on Fusarium oxysporum f. sp. lycopersici associated with Fusarium wilt of tomato. J. of Plant Protec. Res., 50 (2), 175-180.
- [4] Bruce, R.J. and C.A. West (1989). Elicitation of lignin biosynthesis and isoperoxidase activity by pectic fragments in suspension cultures of castor bean. Plant Physiol., 91:889-897.
- [5] Chang, T.H. ; Y.H. Lin ; K.S. Chen ; J.W. Huang ; S.C. Hsiao and P.F. L. Chang (2015). Cell wall reinforcement in watermelon shoot base related to its resistance to Fusarium wilt caused by Fusarium oxysporum f. sp. niveum. J. of Agric. Sci. 153(2), 296-305
- [6] Dane, F. ; L.K. Hawkins ; J.D. Norton ; Y. S. Kwon and Y.H. Om (1998). New resistance to Race 2 of Fusarium oxysporum f.sp. niveum in watermelon. Cucurbit Genet. Coop. Rpt., 21:37-39.
- [7] El-Meghawry, A. ; A. Kamoooh ; M. Abd El-Salam and S.S.Gaman (2001). Inheritance studies of some economic characters in watermelon (Citrullus Lanatus Thunb.) J. Agric. Sci. Mansoura Univ. 26(7): 4159-70.
- [8] Elmstrom, G.W. and D.L. Hopkins. (1981). Resistance of watermelon cultivars to Fusarium wilt. Plant Dis., 65: 825-827.
- [9] El-Shoura, A.M. (2007). Genetic studies on economic traits in watermelon (Citrullus Lanatus Thunb.) Ph.D Thesis, Mansoura Univ. Egypt.
- [10] Esterbaner, H.; E. Schwarzl and M. Hayn (1977). A rapid assay for catechol oxidase and laccase using 2-nitro-5-thio benzoic acid. Anal. Biochem., 77: 486-494.
- [11] Gili, B.S. and J. C. Kumar (1988). Combining ability analysis in watermelon (Citrullus Lanatus Thunb.) Indian J. Hort., 45(2):104-108.
- [12] Gross, G.G. 1980. The biochemistry of lignification. Adv in Botanical Res 8:25-63.
- [13] Hopkins, D.L. and G.W. Elmstrom (1984) . Effect of non-host crop plants on watermelon Fusarium wilt. Plant Dis., 68: 239-241.
- [14] Khereba, A.H.; K. E.A. Abdel-Ati; M.M. Salah and R.R. El-Hawagry (2007). Developing and evaluation of new watermelon hybrids and their heterosis. Egypt. J. Plant Breed., 11(2): 899-923.
- [15] Kosuge, T. (1996). The role of phenolics in host response to infection . Ann. Rev. Phytopathol., 7:195-222.
- [16] Mahdy, A.M.M.; M.H. Abd-El-Mageed; Faten M. Abd-El-Latif; M.M.M. Diab and Nehal M. Saied (2014). Induction of resistance in watermelon plants against Fusarium wilt using chemical inducers and compost under greenhouse conditions. Egypt. J. Phytopathol., 42(2): 1-19
- [17] Massala, R. ; M. Legrand and B. Fritig (1980). Effect of a-aminooxyacetate, a competitive inhibitor of phenylalanine ammonia lyase, on the hypersensitive resistance of tobacco to tobacco mosaic virus. Physiol Plant Pathol., 16: 213-226.
- [18] Martyn, R.D. and R.J.Mclaughlin (1983) . Effects of inoculum concentration on the apparent resistance of watermelons to Fusarium oxysporum f. sp. niveum. Plant Dis., 67: 493-495.
- [19] Martyn, R.D. (1985). Aggressive race of Fusarium oxysporum f.sp. niveum new to the United States. Plant Dis., 69:1007.
- [20] Martyn, R.D. (1987). Fusarium oxysporum f.sp. niveum race 2: A highly aggressive race new to the United States. Plant Dis., 71:233-236.
- [21] Martyn R.D. and D. Netzer. (1991). Resistance to races 0, 1, and 2 of Fusarium wilt of watermelon in Citrullus sp. PI-296341-FR. Hort. Sci., 26: 429-432.
- [22] Netzer, D. (1976). Physiological races of soil population levels of Fusarium wilt of watermelon. Phytoparasitica, 4:131-136.
- [23] Nicholson, R.L. and R. Hammerschmidt (1992). Phenolic compounds and their role in disease resistance. Ann. Rev. of Phytopathol., 30, 369-389.
- [24] Rajan-Bansal; B.S. Sooch and R.K. Dhall 2002. Heterosis in watermelon (Ci-

- trullus Lanatus* Thunb.) .Environmental and Ecol., 20(4):976-979(Computer Res.).
- [25] Simons, T.J. and A.F. Ross (1971). Changes in metabolism associated with enclosed systemic resistance to tobacco. *Phytopathology*, 61:1261-1265
- [26] Sinha, S.K. and R. Khanna (1975). Physiological, biochemical and genetic basis of heterosis. *Advan. in Agron.*, 27(1):123-174
- [27] Snedecor, G.W. and W.G. Cochran (1980). *Statistical Method*, 7th Ed. The Iowa State Univ. Press, Amer., Iowa USA.
- [28] Swiader, M.; M. pronzuk and K. Niemirowicz-Szczytt (2002). Resistance of polish lines and hybrids of watermelon [*Citrullus lanatus* (Thunb.) Matsum et Nakai] to *Fusarium oxysporum* at the seedling stage. *J. Appl. Genet.*, 43(2): 161-170.
- [29] Ulloa, M. ; R.B. Hutmacher ; R.M. Davis ; S.D. Wright ; R. Percy and B. Marsh (2006). Breeding for Fusarium wilt race 4 resistance in cotton under field and greenhouse conditions. *The J. of Cotton Sci.*, 10:114-127
- [30] Wechter, W.P.; C. Kousik ; M. McMillan and A. Levi (2012). Identification of resistance to *Fusarium oxysporum* f. sp. *niveum* race 2 in (*Citrullus lanatus* var. *citroides*) plant introductions. *Hort. Scie.*, 47:334-338.
- [31] Worthington, X.X. (1972). *Enzyme Manual*, Worthington Biochemical Corp., Freehold, New Jersey; 41 -45.
- [32] Zucker, M. (1965). Induction of phenylalanine deaminase by light and its reaction to chlorogenic acid synthesis in potato tuber tissue. *Plant Physiol.*, 40:779-784.

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